

Published on Web 02/11/2006

Modulating the Sensory Response of a Conjugated Polymer by Proteins: An Agglutination Assay for Mercury Ions in Water

Ik-Bum Kim and Uwe H. F. Bunz*

School of Chemistry and Biochemistry, 770 State Street, Georgia Institute of Technology, Atlanta, Georgia 30332.

Received December 12, 2005; E-mail: Uwe.Bunz@chemistry.gatech.edu

Agglutination assays are tools of microbiology and bioanalytical chemistry that identify blood groups, antigen—antibody interactions, prion proteins, and bacteriae such as *S. aureus.*^{1,2} Herein, we describe an agglutination assay for Hg²⁺ as proof of principle for protein-mediated fluorescence modulation of a poly(*para*-phenyleneethynylene) (PPE) **1**. This concept should be broadly applicable and be restricted to neither PPEs nor the proteins we have investigated herein.

Functionalized PPEs are used to detect transition-metal ions, sugar binding proteins (lectins), bacteriae, etc.^{3–6} Schanze et al. and our group have reported upon 1;^{7,8} its solutions are precipitated by the addition of Pb^{2+ 8} but not by Hg²⁺, which leads to weak fluorescence quenching of **1** in PIPES (piperazine-1,4-bis(2-ethanesulfonic acid)) buffer at pH 7.2.

How would other metal ions react with 1? A solution of 1 (5 μ M/repeat unit) in phosphate buffer (0.1 M, pH 7.2)^{8b} was exposed to solutions of 10 different metal ions (0.4 mM). The top panel of Figure 1 shows the results of this experiment when the vials are illuminated by a hand-held UV light ($\lambda_{max} = 365$ nm). At low concentrations of 1, Pb²⁺ ions show slight quenching of the fluorescence of 1 but no precipitation; Zn²⁺, Fe²⁺, and Ni²⁺ also quench the fluorescence of 1, whereas Ca²⁺ ions induce an aggregation induced red shift of the fluorescence of 1.⁹ The other investigated cations (Co²⁺, Cu²⁺, Mg²⁺, Cd²⁺) do not elicit a significant response from 1. Mercury ions (0.4 mM) show the greatest quenching effect on the fluorescence of 1 toward Hg²⁺?

We speculated that the positively charged papain, known to bind to Hg^{2+} , would form an electrostatic complex with 1 in solution. The fluorescence of 1-papain might be sensitive toward Hg²⁺ ions because of the presence of the free sulfhydryl groups in the 1. papain complex.^{8c,10,11d} Proteins modulate the emissive properties and aggregation behavior of 1 and other conjugated polymers.¹² Although we expected papain to be an effective cofactor, we wanted other 1-cofactor complexes (1-histone, 1-bovine serum albumin (BSA), 1-poly(dimethyldiallylammonium chloride) (Pdac, 2)) as controls. Pdac 2 is a positively charged polyelectrolyte. Histone is a compact, positively charged protein used to wrap DNA, and BSA is a negatively charged serum protein used for the biological trafficking of hydrophobic species. We prepared a series of stock solutions where 1 (5 μ M/repeat) was codissolved with Pdac (2) (25 μ M/repeat), histone (5 μ M), BSA (5 μ M), and papain (5 μ M), respectively.

The addition of 2 to 1 leads to a red shift and an increase of fluorescence of 1; 2 wraps around 1, which leads to planarization of the chains and at the same time suppresses excimer formation. The local increase in the refractive index in the complex 1.2 will give an additional red shift. The 1.2 complex is insensitive to the presence of most metal cations; only Fe³⁺ leads to slight quenching. Addition of BSA increases the fluorescence of 1, whereas the fluorescence of 1 is somewhat quenched by histone and papain.^{12a}



Figure 1. Addition of metal ions to PPE 1 in the absence and the presence of cofactors. Blue box: PPE 1 (all samples, $5 \,\mu M$, λ_{max} abs 428 nm). Green box: 1 in the presence of different metals (metal concentration: 0.4 mM). Red box: 1 •cofactor (cofactor concentration: $5 \,\mu M$). Boxed data are controls. The picture shows the fluorescence of different samples under a hand-held UV light (λ_{max} 365 nm). Metal cations are in the +2 oxidation state, with the exception of Fe, which is Fe³⁺.



Figure 2. Left: addition of increasing concentrations of Hg²⁺ (mol/L) to **1** (5 μ M) and its protein and polymer complexes (5 μ M protein). Green box: PPE **1** + increasing concentration of Hg²⁺. Red box: **1**·protein complexes. Blue box: PPE **1** alone. All boxed data are controls. The picture shows the fluorescence of the samples under a hand-held UV light (λ_{max} 365 nm). Right: chemical structures of **1**–**3**.

The cofactors change the response of **1** to a panel of metal cations (Figure 1).¹³ In some cases (see Supporting Information), the highenergy part of the emission is more strongly quenched upon addition of the metal ions; this is probably due to concomitant metal-induced aggregation. The complex of 1-papain gave agglutination with Hg²⁺. Encouraged by these results, we investigated the lowest concentration of Hg^{2+} that led to optical changes of the **1**-cofactor complexes; 1.BSA, 1.histone, and 1.papain were exposed to solutions containing diminishing concentrations of Hg²⁺ ions in water (Figure 2). PPE 1 and 1·BSA are not very sensitive toward Hg²⁺ ions but show visible quenching and/or precipitation at a concentration of 0.1-0.2 mM Hg²⁺. However, the 1-papain complex is already agglutinated in the presence of 20 μ M Hg²⁺ (Figures 2, 3). Although this is not sufficient to detect Hg²⁺ in environmental samples, it is as sensitive as most specifically designed, low molecular weight, mercury-detecting fluorophores.14 How selective is this agglutination assay? Figure 4 shows the results of three experiments. Vial A contains the 1-papain complex. Vial B contains the 1-papain complex in the presence of a mixture of all tested metal cations, and vial C contains 1-papain in the presence of nine metal cations



Figure 3. Qualitative interpretation of the Hg^{2+} -induced agglutination of the 1-papain complex. Top left: 1 alone. Top right: electrostatic complex from 1-papain. Bottom: the addition of Hg^{2+} to 1-papain leads to its precipitation by cross-linking of the papain molecules through Hg^{2+} .



Figure 4. (A) PPE **1**-papain complex (**1**, 5μ M; papain, 5μ M). (B) All 10 metals added to **1**-papain complex (each metal: 0.4 mM). (C) Same without Hg ions. Picture shows fluorescence and was taken under a hand-held UV light (excitation wavelength of 365 nm).

but without Hg^{2+} . The fluorescence of the vials B and C is different, demonstrating that Hg^{2+} ions are *specifically* detected.

Why is the detection of mercury by 1•papain more efficient than by either 1 or papain alone? Figure 3 shows the proposed mechanism of action for this agglutination assay. The complex 1• papain forms as a mixture of oligomers. Positively charged papain molecules are held together by the negatively charged PPE chains. The complex 1•papain *is more sensitive toward agglutination than its parts alone*. The complex will precipitate and leave the supernatant solution nonfluorescent because all of the chains of 1 are incorporated into the Hg²⁺•1•papain agglutinate.

We have found a second system where the same principle can be exploited. The complex of **3**-myosin in phosphate buffer precipitates in 2–5 h. If the test solution is ≥ 0.1 mM of ATP, precipitation is not observed. In the case of myosin itself, only ≥ 1 mM adenosine triphosphate (ATP) prevents myosin from precipitation in phosphate buffer; ATP induces a large conformational change in myosin.¹⁵ The **3**-myosin complex must show a conformation-dependent change in solubility, with the ATP-induced conformation of **3**-myosin being more soluble; this behavior could be used in an ATP sensor.

In conclusion, proteins can modulate the sensory properties of **1** or **3** through electrostatic complex formation. The detection of Hg^{2+} by **1**-papain is proof that specific properties of proteins strongly influence the emissive behavior of conjugated polymers. We will report on further expansion of the concept showing that conjugated polymer—protein complexes will be as powerful as their important and popular DNA hybrids.^{16–18}

Acknowledgment. We thank the Department of Energy, OBES, Division of Material Science and Engineering (Award #DE-FG 0204 ER46141) for generous support and Prof. L. Williams and J. Powers for helpful discussions.

Supporting Information Available: Experimental details, spectroscopic data, and Stern Volmer constants. This material is available free of charge via the Internet at http://pubs.acs.org.

References

- (a) Madigan, M. T.; Martinko, J. M.; Parker, J. Brock Biology of Microorganisms, 10th ed.; Prentice Hall: New York, 2003. (b) Rosi, N. L.; Mirkin, C. A. Chem. Rev. 2005, 105, 1547–1562. (c) Sigal, G. B.; Mammen, M.; Dahmann, G.; Whitesides, G. M. J. Am. Chem. Soc. 1996, 118, 3789–3800. (d) Mortell, K. H.; Weatherman, R. V.; Kiessling, L. L. J. Am. Chem. Soc. 1996, 118, 2297–2298. (e) Arrhenius, S. J. Am. Chem. Soc. 1908, 30, 1382–1388.
- (2) Lee, I. S.; Long, J. R.; Prusiner, S. B.; Safar, J. G. J. Am. Chem. Soc. 2005, 127, 13802–13803.
- (3) Kim, I. B.; Wilson, J. N.; Bunz, U. H. F. Chem. Commun. 2005, 1273– 1275.
- (4) Disney, M. D.; Zheng, J.; Swager, T. M.; Seeberger, P. H. J. Am. Chem. Soc. 2004, 126, 13343–13346.
- (5) Kim, I. B.; Erdogan, B.; Wilson, J. N.; Bunz, U. H. F. Chem.-Eur. J. 2004, 10, 6247-6254.
- (6) Bunz, U. H. F. Adv. Polym. Sci. 2005, 177, 1-52.
- (7) (a) Haskins-Glusac, K.; Pinto, M. R.; Tan, C. Y.; Schanze, K. S. J. Am. Chem. Soc. 2004, 126, 14964–14971. (b) Pinto, M. R.; Schanze, K. S. Synthesis 2002, 1293–1309.
- (8) (a) Kim, I. B.; Dunkhorst, A.; Gilbert, J.; Bunz, U. H. F. *Macromolecules* 2005, 38, 4560–4562. (b) We use phosphate buffer (0.1 M) instead of PIPES buffer here because it is the classic buffer for proteins. (c) Papain, the cystein protease of *Carica papaya*, is commercially used as meat tenderizer and is inexpensive and available in multigram quantities.
- (9) (a) Chen, L. X.; Jager, W. J. H.; Gosztola, D. J.; Niemczyk, M. P.; Wasielewski, M. R. J. Phys. Chem. B 2000, 104, 1950–1960. (b) Wang, B.; Wasielewski, M. R. J. Am. Chem. Soc. 1997, 119, 12–21.
- (10) Smith, E. L.; Kimmel, J. R.; Brown, D. M. J. Biol. Chem. 1954, 207, 533-549.
- (11) (a) Turk, B.; Turk, V.; Turk, D. Biol. Chem. 1997, 378, 141-150. (b) Kimmel, J. R.; Smith, E. L. Adv. Enzymol. 1957, 19, 267-334. (c) Wiederanders, B.; Kaulmann, G.; Schilling, K. Curr. Protein Pept. Sci. 2003, 4, 309-326. (d) Papain: isoelectric point, pH 9.6; dimensions, 5.0 × 3.5 × 3.5 nm and a cleft dissects its long axis. Papain contains seven cystein residues and is 8-fold positively charged at physiological pH and is size commensurate to 1, which is approximately 10-15 nm when extended.
- (12) (a) Kim, I. B.; Dunkhorst, A.; Bunz, U. H. F. *Langmuir* 2005, 21, 7985–7989. (b) Fan, C. H.; Plaxco, K. W.; Heeger, A. J. J. Am. Chem. Soc. 2002, 124, 5642–5643.
- (13) (a) Lavigne, J. J.; Anslyn, E. V. Angew. Chem., Int. Ed. Engl. 1999, 38, 3666–3669. (b) Wiskur, S. L.; Ait-Haddou, H.; Lavigne, J. J.; Anslyn, E. V. Acc. Chem. Res. 2001, 34, 963–972. (c) Rakow, N. A.; Suslick, K. S. Nature 2000, 406, 710–713. (d) Albert, K. J.; Lewis, N. S.; Schauer, C. L.; Sotzing, G. A.; Stitzel, S. E.; Vaid, T. P.; Walt, D. R. Chem. Rev. 2000, 100, 2595–2626.
- (14) (a) Kim, Y.-H.; Youk, S. J.; Moon, S. Y.; Choe, J.-I.; Chang, S.-K. Chem. Lett. 2004, 33, 702-703. (b) Guo, X.; Qian, X.; Jia, L. J. Am. Chem. Soc. 2004, 126, 2272-2273. (c) Nolan, E. M.; Lippard, S. J. J. Am. Chem. Soc. 2003, 125, 14270-14271. (d) Prodi, L.; Bargossi, C.; Montalti, M.; Zuccheroni, N.; Su, N.; Bradshaw, J. S.; Izatt, R. M.; Savage, P. B. J. Am. Chem. Soc. 2000, 122, 6769-6770. (e) Valeur, B.; Leray, I. Coord. Chem. Rev. 2000, 205, 3-40.
- (15) Stryer, L. *Biochemistry*, 4th ed.; W. H. Freeman: New York, 1996; pp 393-396.
- (16) (a) Liu, B.; Bazan, G. C. Proc. Natl. Acad. Sci. 2005, 102, 589–593. (b) Gaylord, B. S.; Heeger, A. J.; Bazan, G. C. J. Am. Chem. Soc. 2003, 125, 896–900. (c) Wang, S.; Bazan, G. C. Adv. Mater. 2003, 15, 1425–1428. (d) Gaylord, B. S.; Heeger, A. J.; Bazan, G. C. Proc. Natl. Acad. Sci. 2002, 99, 12287–12292.
- (17) (a) Chen, L. H.; McBranch, D. W.; Wang, H. L.; Helgeson, R.; Wudl, F.; Whitten, D. G. *Proc. Natl. Acad. Sci.* **1999**, *96*, 12287–12292. (b) Kushon, S. A.; Ley, K. D.; Bradford, K.; Jones, R. M.; McBranch, D.; Whitten, D. *Langmuir* **2002**, *18*, 7245–7249.
- (18) (a) Ho, H. A.; Boissinot, M.; Bergeron, M. G.; Corbeil, G.; Dore, K.; Boudreau, D.; Leclerc, M. Angew. Chem., Int. Ed. Engl. 2002, 41, 1548– 1551. (b) Dore, K.; Dubus, S.; Ho, H. A.; Levesque, I.; Brunette, M.; Corbeil, G.; Boissinot, M.; Boivin, G.; Bergeron, M. G.; Boudreau, D.; Leclerc, M. J. Am. Chem. Soc. 2004, 126, 4240–4244.

JA058431A